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Microbiological and chemical characteristics of *pasta filata* type cheese from raw ewe milk, using thermophilic and mesophilic starters

Mikrobiologische und chemische Eigenschaften von Käse vom Typ „Pasta filata“ aus roher Schafsmilch unter Verwendung von thermophilen und mesophilen Startern

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Summary

The aims of this study were a) to produce *mozzarella* type cheese from raw ewe milk at both high and low temperatures; b) to verify the effect of vacuum packaging on the microbiological characteristics of a low moisture *pasta filata* cheese. In the colder season, the use of both thermophilic and mesophilic starters allowed the reduction of the curd acidification time as compared to the use of thermophilic starters. The microbiological characteristics were not affected by the different acidification times. Pathogens and hygiene markers microorganisms were undetected in both group of *mozzarella* cheese. No differences were found for *Enterobacteriaceae* and coliform counts. Instead, the proteolysis marker was higher at a longer curd acidification. The decreasing of microorganisms used as hygiene markers as well as *Enterobacteriaceae* and coliforms present in milk and curd was observed in both groups of *mozzarella* cheese. It could be a result of the thermal effect of the stretching phase. Low moisture *pasta filata* cheese upon storage under vacuum for 40 days, according to microbiological results, is free of risks for consumer health.

Keywords: raw ewe milk, mozzarella, starters, vacuum storage

Zusammenfassung

Die Ziele dieser Studie waren: a) Herstellung von Mozzarella-Käse aus roher Schafsmilch bei hohen und niedrigen Temperaturen; b) Überprüfung der Wirkung von Vakuumverpackungen auf mikrobiologische Eigenschaften. In kälteren Jahreszeiten erlaubte die Verwendung von sowohl thermophilen als auch mesophilen Startern eine Verkürzung der Säuerungsdauer im Vergleich zu der Verwendung von thermophilen Startern. Mikrobiologische Eigenschaften wurden durch unterschiedliche Ansäuerungszeiten nicht beeinflusst. Pathogene und Hygienemarker Mikroorganismen wurden in beiden Mozzarella-Käsegruppen nicht entdeckt. Für Enterobacteriaceae und coliforme Zählungen wurden keine Unterschiede gefunden. Es ergab sich dagegen, dass der Proteolysemarker höher ist, wenn die Ansäuerung des Quarks länger ist. Die Abnahme von Mikroorganismen, die als Hygienemarker verwendet werden, sowie von Enterobacteriaceae und Coliformen, die in Milch und Quark enthalten sind, wurde in beiden Gruppen von Mozzarella-Käse beobachtet. Es könnte ein Ergebnis der thermischen Wirkung der Dehnungsphase sein. Pasta-Filata-Käse mit niedriger Feuchtigkeit, der nach 40-tägiger Lagerung unter Vakuum aufbewahrt wird, ist gemäß den mikrobiologischen Ergebnissen frei von Risiken für die Gesundheit der Verbraucher.

Schlüsselwörter: rohe Schafsmilch, *Mozzarella*, Startern, Vakuumverpackung

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Introduction

Sheep production is widespread in the Mediterranean area, including the Center as well as the South of Italy, and is mainly oriented to milk production. Most of the milk is used to produce hard and semi-hard cheeses. Some cheese types are famous and recognized as Protected Designation of Origin (PDO) by the European Union: *Pecorino Romano*, *Pecorino Sardo*, *Canestrato Pugliese*, *Idiazabal*, *Manchego* and *Roquefort* (Scintu and Piredda, 2007). Recently, also *Vastedda della Valle del Belice*, made only from ewe milk, was recognized as PDO cheese. It is a so called *pasta filata* cheese, though in Italy, this type of cheese is mainly produced from cow and buffalo milk. Ewe milk, mixed with cow and goat milk, is also used in the production of *Kashkaval*, another *pasta filata* type cheese, very popular in Eastern Europe. *Pasta filata* cheese is characterized by the stretching phase, which follows the curd acidification. The stretching phase consists in submitting curd to elongation in presence of hot water (90–95 °C). Casein aggregates are bound and form long chains, which entraps fat and moisture (Addeo et al., 1996). The poor stretching ability of ewe milk (Niro et al., 2014) is probably the main limit to the spread of *pasta filata* cheese type from ewe milk. Recently, increased interest for “*pasta filata*” cheese was made evident also for ewe milk. The cheese making process used with cow milk to produce *Scamorza* was transferred to ewe milk (Albenzio et al., 2013a; Albenzio et al., 2013b). Ewe milk was inoculated with starters and at pH of 5.7 liquid rennet was added. Indeed, the trend and the length of the acidification phase depend on the temperature that affects the growth of lactic acid bacteria (LAB). In small cheese plants not oriented to *pasta filata* type and not equipped with temperature-controlled vessels, it is more difficult to manage the temperature during curd acidification. In fact, the trend and the length of the acidification phase depend on the temperature that affects the growth of lactic acid bacteria (LAB). One of the methods used to improve the management of the acidification phase is the addition of starter cultures, which can grow at different temperatures, both at high temperatures during summer as well as at low temperatures during winter (Gaglio et al., 2014). According to Faccia et al. (2015), the most appropriate technology to produce soft *pasta filata* cheese from ewe and goat milk in small dairies is a combination of direct acidification and lactic fermentation.

The present work was developed to study how improve *pasta filata* cheese technology in small cheese plants which process ewe milk. Regarding cheese packaging, the effects of vacuum packaging, the most common packaging system used in small cheese plants, was evaluated only on ripened cheese (Nuñez et al., 1986; AGRIS Sardegna, 2011; Salari et al., 2011). Therefore, studies on soft cheese are useful to evaluate the effect of the vacuum packaging on microbiological characteristics.

The aims of this study are therefore to evaluate the microbiological and chemical characteristics of a) *mozzarella* cheese produced from raw ewe milk, with inoculation of different types of starters and b) low-moisture *pasta filata* cheese during storage by vacuum packaging.

Materials and methods

Process

Mozzarella type cheese. The trial was carried out in a small cheese plant in autumn 2013. Four replicates per *mozzarel-*

la processing were performed. Regarding to the Brucellosis disease, Lazio region was declared free from the infection of *Brucella melitensis* (Decision 2014/91 UE). Raw ewe milk (100 l for each replicate) was inoculated with two kinds of commercial starters. Starter 1: thermophilic (T) lactic acid bacteria (LAB) containing only *Streptococcus thermophilus* (STA IDC 11; Centro Sperimentale del Latte SpA, Vernate, Milano). Starter 2: thermophilic and mesophilic (TM) lactic acid bacteria (LAB) with *Streptococcus thermophilus*, *Lactococcus lactis*, *L. cremoris* and *L. diacetylactis* (STAR IDC 12; Centro Sperimentale Latte, Vernate, Milano). After the starter inoculation, milk was added with rennet powder (Bellucci S.R.L., Modena, Italy). After coagulation, curd was cut and let acidify under whey. The vessel containing curd under whey was put in handmade bath in order to maintain curd temperature not below 37 °C. In order to verify whether acidified curd was ready to be stretched, a “stretching” test was carried out on a piece of curd treated with hot water. At the same time, the pH of curd was measured. During the stretching phase, hot water at about 85–90 °C was used. Cheese was then molded in shape of about 200 g, firmed in cold water, salted in saturated brine for 2 hours and stored at 4 °C into governing liquid with 2–3 % of NaCl.

Low-moisture *pasta filata* cheese. Another *pasta filata* cheese was created in order to study the effect of storage by vacuum packaging. This product weighted about 50 g. The process was similar to the one described above and included TM starter. After salting, cheese was left at 4 °C for some hours to reduce the moisture content. Then, it was wrapped in a special paper (Arcadia SpA, Sedegliano, Udine) and put under vacuum. This paper adheres perfectly to the cheese, so reducing air and humidity areas that, during storage under vacuum, could favor the growth of moulds. For the vacuum packaging, an envelope-embossed polyamide/polyethylene has average gas barrier.

Microbiological and chemical analyses

Mozzarella type cheese. During the four replicates, milk and curd samples were collected and submitted to microbiological and chemical analyses. All microbiological analyses were performed in the laboratories of IZSLT (Istituto Zooprofilattico Sperimentale Lazio e Toscana – Experimental Institute for Zooprophyllaxis in Lazio and Tuscany), accredited according to International Organization for Standardization (ISO/IEC 17025:2005) by Accredia, (Laboratory number 0201), the Italian Accreditation Body. Samples of T and TM cheese were collected at day 1 (the day after processing) and day 5. Samples of milk, curd and cheese of day 1 were submitted to the following microbiological analyses: detection of *Salmonella* (ISO 6579:2002), and *Listeria monocytogenes* (ISO 11290-2:1998/Amd 1:2004), enumeration of coagulase-positive staphylococci (ISO 6888-2:1999/Amd 1:2003), β -glucuronidase-positive *E. coli* (ISO 16649-2: 2001), *Enterobacteriaceae* (ISO 21528-2:2004), coliforms at 30 °C (ISO 4832: 2006) and total microbial count (TMC) at 30 °C (ISO 4833-2:2013), (Filipetti et al., 2008; Carfora et al., 2013). At day 5, coliforms at 30 °C analyses were performed on cheese samples.

The following chemical and technological analyses were carried out on milk samples: total nitrogen (TN) and fat by NIR (Milkoscan FT 6000 – Foss Electric, Hillerød, Denmark) calibrated for ewe milk; pH was measured with pH meter (Mettler Toledo Inc., Leicester, UK). Clotting properties (r = rennet clotting time, K20 = curd firming

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time, A30 = curd firmness at 30 min, A2r = curd firmness at 2r) by Zannoni and Annibaldi method (1981).

Cheese samples collected at day 1 and day 5 were submitted to the following chemical analyses: pH, moisture (IDF, 1986), total nitrogen (TN) (FIL-IDF, 1993), soluble nitrogen (SN) (FIL-IDF, 1991), fat (FIL-IDF, 2001), salt (IDF, 1988).

Low-moisture *pasta filata* cheese. Samples stored by vacuum packaging were collected at 0, 20 and 40 days of storage and submitted to the following analyses: *Salmonella*, *Listeria monocytogenes*, coagulase-positive staphylococci, β -glucuronidase-positive *E. coli*, *Enterobacteriaceae*, coliforms at 30 °C, TMC, yeasts and moulds (ISO 21527-1: 2008), (Filippetti et al., 2008; Carfora et al., 2013). Moisture, protein and fat content were analyzed by NIR spectroscopy (FoodScan™, Foss Electric, Hillerød, Denmark) (Filippetti et al., 2008).

Statistical analysis

All the microbiological data were log-transformed before statistical analysis. Samples showing no colonies on plates counts were converted to zero after log-transformation. The GLM procedure of SAS software (SAS Institute Inc. 2007) was used for statistical analysis of microbiological and chemical analyses of *mozzarella* type cheese. Factorial model including the fixed effect of the starter type and the storage time of cheese was used.

Results and discussion

Mozzarella type cheese – Process

During the four replicates carried out on *Mozzarella* cheese, both T and TM curds were successfully stretched at pH 5.0. The length of the acidification phase was affected by the kind of starter. In fact, TM curd was ready for stretching in four hours and a half on average, while T curd in seven hours on average. Curd acidification begins when the whey is drained off (McSweeney et al., 2004). At the conditions of the present trial (autumn), the use of starters based on the thermophilic and mesophilic LAB allowed to reach an optimal curd pH in about the same time currently required in the cheese-making of buffalo and cow *mozzarella*, 3–5 hours starting from whey draining (Mucchetti and Neviani, 2006). The necessary time to reach optimal curd pH depends mainly on curd temperature, recommended to be not less than 30 °C (Addeo et al., 1996; Salvadori del Prato, 2001). In the Mediterranean countries, ewe milk is produced also during cold seasons, when the management of the acidification phase is difficult due to the lack of specialized facilities in traditional or small cheese plants. The use of starters containing both thermophilic and mesophilic LAB could favor the acidification at different temperatures and allow to carry out the acidification phase in a reasonable time. *Vastedda della Valle del Belice* cheese was traditionally produced only in summer with raw milk and with no starters, but it is currently requested throughout the year. Recently, twelve thermophilic and mesophilic LAB strains isolated from PDO *Vastedda della Valle del Belice* cheese were studied for the full-year production of *Vastedda*-like cheese with raw milk and natural

starters (Gaglio et al., 2014). In *Fior di Latte* from raw sheep milk processed by the direct acidification and lactic fermentation, time from whey draining to stretching was about 5 hours (Faccia et al., 2015).

Mozzarella type cheese – Microbiological characteristics

Table 1 displays the microbiological characteristics of milk, curd and *mozzarella* type cheese at day 1 and day 5.

Salmonella and *L. monocytogenes*, pathogens considered as food safety criteria (Reg CE 2073/2005), were undetected by qualitative analyses in milk, curd and cheese, in accordance with the above Regulation.

In the cheese samples, counts of microorganisms considered as hygiene markers (Reg CE 2073/2005), β -glucuronidase-positive *E. coli* and coagulase-positive staphylococci were lower than the detection limit of the method. According to Reg CE 2073/2005 the detection of staphylococcal enterotoxins was not carried out due to the absence of coagulase-positive staphylococci.

Higher counts of these microorganisms were found in milk and curd. The average coagulase-positive staphylococci in milk and curd samples, was, respectively, 2.0 log 10 cfu x g⁻¹ and 2.4 log 10 cfu x g⁻¹. These data are in accordance with regulatory limits and, according to Italian legislation, the detection of staphylococcal enterotoxins was not carried out. The average β -Glucuronidase-positive *E. coli* in milk and curd samples, was, respectively, 1.8 log 10 cfu x g⁻¹ and 3.3 log 10 cfu x g⁻¹.

In another *pasta filata* fresh cheese from raw milk as *Vastedda della Valle del Belice* coagulase-positive staphylococci and β -Glucuronidase-positive *E. coli* counts resulted positive (Mucchetti et al., 2008). The same microorganisms were found to be lower than the detection limit of the method on *Mozzarella di Bufala Campana* from pasteurized milk, while on the curd their number was very variable according to the different trials (Zottola et al., 2009). In *Fior di Latte* from raw sheep milk, coagulase-positive staphylococci were detected (Faccia et al., 2015).

Enterobacteriaceae and coliform counts were positive in cheese samples. The average of *Enterobacteriaceae* resulted 1.7 log 10 cfu x g⁻¹ in T *mozzarella* type cheese and 1.3 log 10 cfu x g⁻¹ in TM ones. The average coliform was 1.5 log 10 cfu x g⁻¹ in T *mozzarella* like cheese and 1.4 log

TABLE 1: Microbiological characteristics of milk, curd and *mozzarella* type cheese at day 1 and 5.

	Milk	Curd	day 1 mozzarella T	day 1 mozzarella TM	day 5 mozzarella T	day 5 mozzarella TM
<i>Salmonella</i> spp. (in 25 g)	absent	absent	absent	absent	-	-
<i>Listeria monocytogenes</i> (in 25 g)	absent	absent	absent	absent	absent	absent
Coagulase positive staphylococci (log 10 cfu x g ⁻¹)	2.0	2.4	0.0 (*)	0.0 (*)	-	-
Beta-glucuronidase-positive <i>E. coli</i> (log 10 cfu x g ⁻¹)	1.8	3.3	0.0 (*)	0.0 (*)	-	-
<i>Enterobacteriaceae</i> (log 10 cfu x g ⁻¹)	4.9	5.2	1.7	1.30	-	-
Coliforms (log 10 cfu x g ⁻¹)	5.0	5.1	1.5	1.4	2.2	1.3
Total mesophilic count (log 10 cfu x g ⁻¹)	6.1	7.1	6.6	6.9	7.5	7.6

T: thermophilic lactic acid bacteria; TM: thermophilic and mesophilic lactic acid bacteria; (*) < 10 cfu x g⁻¹

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10 cfu x g⁻¹ in TM ones. Significant differences between T and TM cheese were not found. *Enterobacteriaceae* found on Vastedda fresh cheese (6.3 log₁₀ cfu x g⁻¹) was higher if compared with our results (Mucchetti et al., 2008).

The average TMC increased from milk (6.1 log₁₀ cfu x g⁻¹) to curd (7.1 log₁₀ cfu x g⁻¹) and slightly decreased in T (6.6 log₁₀ cfu x g⁻¹) and TM cheese (6.9 log₁₀ cfu x g⁻¹). Regulatory limits are present only for raw milk, Regulation (EC) 853/2004, and are respected by samples. Table 3 displays microbiological characteristics of *mozzarella* cheese after 5 days of storage. The search for *L. monocytogenes* was negative. The average coliform count was 2.2 log₁₀ cfu x g⁻¹ in T *mozzarella* like cheese and 1.4 log₁₀ cfu x g⁻¹ in TM ones. The average TMC was 7.5 log₁₀ cfu x g⁻¹ in T *mozzarella* type cheese and 7.6 log₁₀ cfu x g⁻¹ in TM ones. These values of TMC could be attributed to starter cultures (Gaglio et al., 2014). No significant differences in coliforms were found between T and TM cheese. However, it is noteworthy that the coliforms in T cheese increased by 48 % from day 1 to day 5, while the values found in TM cheese were very similar (Fig. 1).

The mean values of coliforms in all cheese (T and TM), collected at day 1 and 5, were compared with milk and curd (Fig. 2). Coliform count, similar in milk and curd, decreased by 72 % from curd to fresh cheese and increased during cheese storage (19 %). Similar decreasing from curd to fresh cheese was observed for *Enterobacteriaceae* (71 %).

Raw milk cheese deserve greater attention since the final products can become contaminated by pathogenic microorganisms as a result of their presence in raw milk (Donnelly, 2004). The growth of milk pathogens in raw cheeses is highly dependent on the variety of cheese and on the involved technology. It is well documented that pathogens grow more easily in cheese with higher moisture, high pH and low salt content (Grappin and Beuvier, 1997).

The decreasing of microorganisms used as hygiene markers as well as *Enterobacteriaceae* and coliforms from milk and curd to cheese could be a result of the thermal effect of the stretching phase. The high temperatures reached during this phase of *pasta filata* processing contributes to reduce contaminating microbial flora and to the safety of the resulting products (Mucchetti and Neviani, 2006).

Based on all microbiological results, the higher acidification time of T cheese did not result in poor hygienic characteristics, also during cheese storage. During the trial, the applied process contributed to reduce the microbiological risk of cheese produced from raw milk. According to Johnson et al. (1990), the good management of curd acidification can contribute to reducing microbiological risk and/or increasing shelf life of cheese from raw milk. In the small cheese plants not equipped with temperature-controlled vessels, curd acidification could be managed by an appropriate use of starters (Niro et al., 2014, Settanni et al., 2012).

Mozzarella type cheese – Physico-chemical characteristics

The mean physical, chemical and technological characteristics of the milk used in *mozzarella* type cheese trials

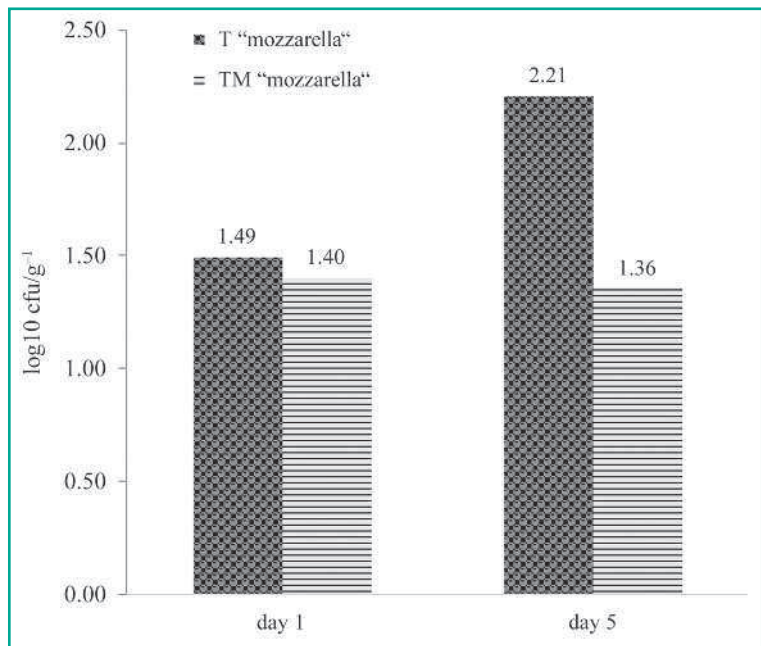


FIGURE 1: Coliform count of T and TM "mozzarella" cheese at day 1 and 5.

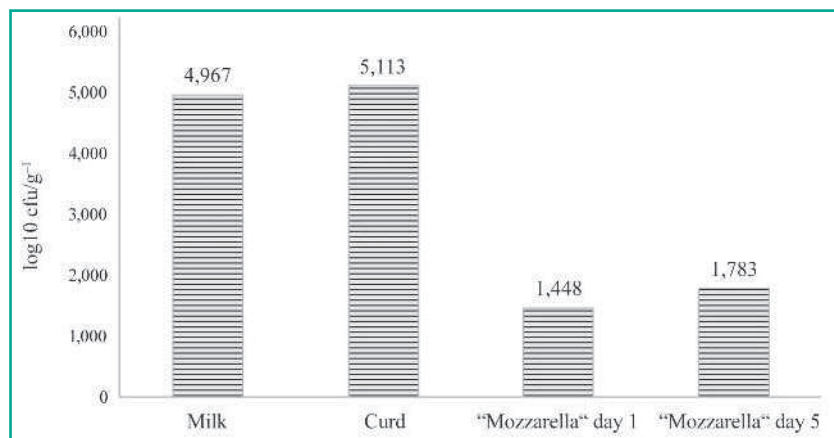


FIGURE 2: Coliform count of milk, curd and "mozzarella" cheese (T and TM type) at day 1 and 5.

were as follows: pH 6.72, total protein 5.64 %, fat 6.33 %, r 12.97 min, k20 8.25 min and A30 28.44 mm.

The chemical characteristics of the T and TM *mozzarella* type cheese are reported in Table 2. At day 1, the moisture of the T cheese was significantly lower ($P < 0.05$) than the TM one (58.12 vs 61.27 %). The lower moisture of T *mozzarella* type cheese could be due to the longer acidification time, in which the higher rate of curd syneresis occurred. Rapid acidification allows the total manufacturing time to be shortened, because it reduces the total amount of syneresis during cheese making and enables a higher moisture content to be achieved in the final cheese (Barbano et al., 1994).

The mean moisture content of both T and TM cheese at day 1 was 59.69 % and was slightly higher than the one of *mozzarella* from cow and buffalo milk, 58.8 and 55.5 %, respectively¹⁾ (1). Both *Vastedda della Valle del Belice* and *Scamorza* cheese have lower moisture content than the *mozzarella* type cheese, respectively, 46.7 % (Mucchetti et al., 2008) and 44.9 % (nut.entecra.it/646/tabelle_di_compo-

¹⁾ nut.entecra.it/646/tabelle_di_composizione_degli_alimenti.html, 2009

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sizione_degli_alimenti.html, 2009) and they are included in the semi-hard cheese category of Codex Alimentarius. The *Mozzarella* type cheese produced in the present trial could be ascribed to the soft cheese category of Codex Alimentarius. In spite of the scarce ability of ewe milk to be stretched, it was possible, to obtain a product similar to the *mozzarella* obtained from cow and buffalo milk through bacterial acidification,

The moisture content of day 5 samples was higher on average than the one of day 1 cheese. Instead, salt content and, consequently, ash content, of whom NaCl represents the highest component, decreased from day 1 to day 5. During storage in governing liquid, cheese absorbed water while releasing salt into the governing liquid.

The values of the soluble protein showed that protein degradation was higher in the T cheese especially at day 5 (1.55 vs 1.13 %) ($P < 0.05$). *Mozzarella* is a fresh cheese where the extent of proteolysis is very limited (Upadhyay et al., 2004). Proteolysis during cheese aging is a well-known biochemical reaction (Upadhyay et al., 2004), but it remains less known in *pasta filata* cheese such as *mozzarella* (Johnson and Law, 1999; Feeney et al., 2002). Moisture content generally affects the extent of proteolysis (Upadhyay et al., 2004). Out of the factors, which mostly affect *mozzarella* proteolysis, lactic acid bacteria have to be focused (Barbano et al., 1993; Hutkins, 1993; Feeney et al., 2002). The process type and management of (Hynes et al., 2004) affect both the moisture content and lactic acid bacteria directly or indirectly. Some undesirable changes in the characteristics of *mozzarella* can be due to casein proteolysis (Rudan et al., 1999; Feeney et al., 2002). We can hypothesize that, in the trial conditions, a longer curd acidification could favor the proteolytic degradation.

The pH values of both kind of *mozzarella* decreased after 5 days of storage.

Low-moisture *pasta filata* cheese – Microbiological and physico-chemical characteristics

Table 3 displays the microbiological characteristics of curd and low-moisture *pasta filata* cheese at day 0, 20 and 40. *Salmonella* and *L. monocytogenes* were both absent in 25 g. β -Glucuronidase-positive *E. coli*, coagulase-positive staphylococci, *Enterobacteriaceae* and coliforms were detected in curd but not in cheese at 0, 20 and 40 days, where they were lower than the detection limit of the method. The mean total mesophilic count was lower in curd ($7.0 \log_{10} \text{ cfu} \times \text{g}^{-1}$) and increased during storage of cheese: day 0 ($8.3 \log_{10} \text{ cfu} \times \text{g}^{-1}$), day 20 ($8.4 \log_{10} \text{ cfu} \times \text{g}^{-1}$) and day 40 ($9.1 \log_{10} \text{ cfu} \times \text{g}^{-1}$). Yeasts, higher in curd, were lower than the detection limit of the method at day 0 and grew during cheese storage. Mould content, present in

TABLE 2: Physico-chemical characteristics of *mozzarella* type cheese at day 1 and 5.

	day 1		day 5	
	T	TM	T	TM
moisture (%)	58.12 ^b	61.27 ^a	64.62	62.44
protein (%)	20.66	18.31	16.40	17.11
soluble protein (%)	1.38	1.26	1.55 ^a	1.13 ^b
fat (%)	17.23	15.89	15.87	15.34
NaCl (%)	0.80	0.77	0.60	0.67
pH	4.99	5.00	4.91	4.91

T: thermophilic lactic acid bacteria; TM: thermophilic and mesophilic lactic acid bacteria, ^{a, b}: $P < 0.05$

TABLE 3: Microbiological characteristics of snack cheese at day 0, 20 and 40.

	Curd	Low-moisture <i>pasta filata</i> cheese		
		day 0	day 20	day 40
<i>Salmonella</i> spp. (in 25 g)	absent	absent	absent	absent
<i>Listeria monocytogenes</i> (in 25 g)	absent	absent	absent	absent
Coagulase positive staphylococci ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	3.1	0.0(*)	0.0(*)	0.0(*)
Beta-glucuronidase-positive <i>E. coli</i> ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	2.3	0.0(*)	0.0(*)	0.0(*)
Enterobacteriaceae ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	5.6	0.0(*)	0.0(*)	0.0(*)
Coliforms ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	2.5	0.0(*)	0.0(*)	0.0(*)
Total mesophilic count ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	7.0	8.2	8.4	9.1
Yeasts ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	5.5	0.0(*)	3.3	2.9
Moulds ($\log_{10} \text{ cfu} \times \text{g}^{-1}$)	4.0	0.0(*)	2.8	0.0(*)

(*) $< 10 \text{ cfu} \times \text{g}^{-1}$

curd, was lower than the detection limit of the method at day 0 and day 40, detected at day 20.

The chemical analyses of the low-moisture *pasta filata* cheese, showed that the mean moisture content was 50.00 %, protein content 24.20 % and fat content 22.33 %, corresponding to 298 kcal/100 g of product and 149 kcal per portion.

Microbiological results of fresh cheese were satisfactory, if compared with those reported by other Authors. In fresh *Vastedda della Valle del Belice*, having the similar moisture content (53.30 %), coagulase-positive staphylococci, β -Glucuronidase-positive *E. coli* and *Enterobacteriaceae* were always present (Mucchetti et al., 2007). The coliforms found in the *mozzarella* cheese stored under vacuum (Tanweer and Goyal, 2011) were always higher than $2.0 \log_{10} \text{ cfu} \times \text{g}^{-1}$. Coliforms, *E. coli*, mould and yeasts were detected from day 1 to day 24, in *mozzarella* cheese, having about 50 % of moisture content and stored under vacuum (Fuentes et al., 2015).

The data collected in our trials showed that the storage of low-moisture *pasta filata* cheese at 4 °C, under vacuum, for 40 days is free of risks for the consumer health, according to the considered parameters. Indeed, the vacuum packaging resulted the best method to store *Vastedda della Valle del Belice*, if compared to the other packaging systems (Palmeri et al., 2015).

Conclusions

To satisfy the market demand of new cheese types from ewe milk, and especillay fresh cheese, it is necessary to develop new cheese processes. The *pasta filata* cheese, like *mozzarella*, is a kind of cheese, which is highly requested by consumers, but, up to now, the poor stretching ability of ewe milk has limited the application of this process with ewe milk. The use of a mix of microorganisms able to grow at different temperatures could favor curd acidification so reducing the acidification time in the colde season. The use

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of both thermophilic and mesophilic starters, in experimental trials, allowed to reduce the curd acidification time compared to the use of thermophilic starters. Based on the microbiological results obtained, the longer acidification time of the cheese inoculated with the thermophilic lactic acid bacteria did not result in poor hygienic characteristics. The microorganisms used as hygiene markers (*E. coli* and coagulase positive staphylococci) were not detectable in both kinds of fresh cheese. In the same way, *Enterobacteriaceae* and coliform counts were analogous in the two kinds of fresh cheese. Even after a few days of storage, the coliform count did not differ between the two kinds of cheese. The use of different starters affected some chemical characteristics: in the fresh cheese, in the products with a longer acidification time, the moisture content was lower, while, during storage, the proteolytic process was favoured. This could imply some undesirable characteristics of *mozzarella* cheese.

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The results of this work could be useful to promote technological innovation in small cheese plants processing ewe milk.

Conflict of interest

The authors declare that there is no conflict of interest.

References

- Addeo F, Emaldi GC, Masi P (1996): Tradition and innovation in the “Mozzarella di Bufala Campana cheese” production. In: EAAP (ed.), International symposium on buffalo products. Wageningen Pers, Wageningen, ND, 23–39.
- AGRIS Sardegna, LAORE, Università degli Studi di Sassari (2011): Stima della shelf-life di formaggi ovini per l'applicazione di procedure operative impiegabili dalle aziende del comparto. LAORE Sardegna, Cagliari, IT.
- Albenzio M, Santillo A, Caroprese M, Ruggieri D, Napolitano F, Sevi A (2013a): Physicochemical properties of Scamorza ewe milk cheese manufactured with different probiotic cultures. *J Dairy Sci* 96: 2781–2791.
- Albenzio M, Santillo A, Caroprese M, Braghieri A, Sevi A, Napolitano F (2013b): Composition and sensory profiling of probiotic Scamorza ewe milk cheese. *J Dairy Sci* 96: 2792–2800.
- Barbano DM, Chu KY, Yuri JJ, Kindstedt PS (1993): Contribution of coagulant, starter and milk enzymes to proteolysis and browning in Mozzarella cheese. In: Proc. 30th Marshall Italian and Specialty Cheese Seminars, Rhodia-Marshall, Madison, USA, 65–79.
- Barbano DM, Yun JJ, Kindstedt PS (1994): Mozzarella cheese making by a stirred curd, no brine procedure. *J Dairy Sci* 77: 2687–2694.
- Carfora V, Marri N, Tripaldi C, Patriarca D, Giacinti G, Boselli C, Giangolini G, Amatiste S (2013): Yoghurt from short supply chain: preliminary study of microbiological and physicochemical characteristics during shelf life. *Ital J Food Safety* 2: 14–16.
- Decision nr. 2014/91/UE, 14.02.2014 Commission Implementing Decision, 14 february 2014, amending Annex II to the Decision 93/52/CEE, recognizing as officially free of brucellosis (*B. melitensis*) some regions of Italy and Spain, and Annexes I, II and III to the Decision 2003/467/CE as regards the declaration that Hungary is officially tuberculosis-free, Romania and some regions of Italy are officially free of brucellosis and some regions of Italy are officially free of enzootic bovine leucosis.
- Donnelly CW (2004): Growth and survival of microbial pathogens in cheese. In: Fox PF, McSweeney PLH, Cogan TM, Guinee TP (ed.) “Cheese: Chemistry, physics and microbiology, vol 1 General aspects, Elsevier, London, 541–559.
- Gaglio R, Scatassa ML, Cruciatà M, Miraglia V, Corona O, Di Gerlando R, Portolano B, Moschetti G, Settanni L (2014): In vivo application and dynamics of lactic acid bacteria for the four-season production of Vastedda-like cheese. *Int J Food Microbiol* 177: 37–48.
- Faccia M, Trani A, Gambacorta G, Loizzo P, Cassone A, Caponio F (2015): Production technology and characterization of Fior di latte cheeses made from sheep and goat milks. *J Dairy Sci* 98: 1402–1410.
- Feeney EP, Guinee TP, Fox PF (2002): Effect of pH and calcium content on proteolysis in Mozzarella cheese. *J Dairy Sci* 85: 1646–1654.
- FIL-IDF (1991): Chemical methods for evaluating proteolysis in cheese maturation, No. 261. Brussels, Belgium.
- FIL-IDF (1993): Milk. Determination of nitrogen content. Kjeldahl method; Standard 20B. Annex I: Modified procedure for milk products. Brussels, Belgium.
- FIL-IDF 172 (2001)/ISO 14156 (2001): Milk and Milk products-Extraction methods for lipids and liposoluble compounds. Brussels, Belgium.
- Filippetti F, Giangolini G, Boselli C, Amatiste S, Patriarca D, Rosati R (2008): Ricotta Romana: manufacturing technique, chemical and microbiological composition. In: Proc. FIL IDF World Dairy Summit & Exhibition, 11–14 November 2008, Mexico City, Mexico.
- Fuentes L, Mateo J, Quinto EJ, Caro I (2015): Changes in quality of nonaged pasta filata Mexican cheese during refrigerated vacuum storage. *J Dairy Sci* 98: 2833–2842.
- Grappin R, Beuvier E (1997): Possible implications of milk pasteurization on the manufacture and sensory quality of ripened cheese. *Int Dairy J* 7: 751–761.
- Hynes ER, Aparo L, Candiotti MC (2004): Influence of residual milk-clotting enzyme on α s1 casein hydrolysis during ripening of Reggiano Argentinian cheese. *J Dairy Sci* 87: 565–573.
- http://nut.entecra.it/646/tabelle_di_composizione_degli_alimenti.html
- Hutkins RW (1993): How starter culture metabolism affects properties of Mozzarella cheese. In: Proc. 30th Marshall Italian and Specialty Cheese Seminars, Rhodia-Marshall, Madison, USA, 1–80.
- IDF (1986): Cheese and processed cheese products. Determination of dry matter. IDF Standard No. 4. FIL-IDF, Brussels, Belgium.
- IDF (1988): Determination of dry salt content. IDF Standard No. 12B. FIL-IDF, Brussels, Belgium.
- ISO, 1998b /2004: Microbiology of food and animal feeding stuffs – Horizontal method for the detection and enumeration of *Listeria monocytogenes* – Part 2: Enumeration method. ISO regulation 11290-2:1998/Amd 1:2004. International Standard Organization, Geneva, Switzerland.
- ISO, 1999b /2003: Microbiology of food and animal feeding stuffs – Horizontal method for the enumeration of coagulase-positive staphylococci (*Staphylococcus aureus* and other species) – Part 2: Technique using rabbit plasma fibrinogen agar medium. ISO regulation 6888-2:1999/Amd 1:2003. International Standard Organization, Geneva, Switzerland.

The contents are protected by copyright. The distribution by unauthorized third parties is prohibited.

- ISO, 2001b:** Microbiology of food and animal feeding stuffs – Horizontal method for the enumeration of beta-glucuronidase-positive *Escherichia coli* – Part 2: Colony-count technique at 44 degrees C using 5-bromo-4-chloro-3-indolyl beta-D-glucuronide. ISO regulation 16649-2:2001. International Standard Organization, Geneva, Switzerland
- ISO, 2002:** Microbiology of food and animal feeding stuffs – Horizontal method for the detection of *Salmonella* spp. ISO regulation 6579:2002. International Standard Organization, Geneva, Switzerland.
- ISO, 2004b:** Microbiology of the food chain – Horizontal method for the detection and enumeration of Enterobacteriaceae – Part 2: Colony-count technique. ISO regulation 21528-2:2004. International Standard Organization, Geneva, Switzerland.
- ISO, 2006:** Microbiology of food and animal feeding stuffs – Horizontal method for the enumeration of coliforms – Colony-count technique. ISO regulation 4832:2006. International Standard Organization, Geneva, Switzerland.
- ISO, 2008a:** Microbiology of food and animal feeding stuffs – Horizontal method for the enumeration of yeasts and moulds – Part 1: Colony count technique in products with water activity greater than 0.95. ISO regulation 21527-1:2008. International Standard Organization, Geneva, Switzerland.
- ISO, 2013b:** Microbiology of the food chain – Horizontal method for the enumeration of microorganisms – Part 2: Colony count at 30 degrees C by the surface plating technique. ISO regulation 4833-2:2013. International Standard Organization, Geneva, Switzerland.
- ISO/IEC, 2005:** General requirements for the competence of testing and calibration laboratories. ISO regulation 17025:2005. International Standard Organization, Geneva, Switzerland.
- Italian National Accreditation Body** <http://www.accredia.it/> [Last access 13th April 2018].
- Johnson EA, Nelson JH, Johnson M (1990):** Microbiological Safety of Cheese Made from Heat-Treated Milk, Part II. *Microb. J Food Prot* 53: 519–540.
- Johnson M, Law BA (1999):** The origins, development and basic operations of cheesemaking technology. In: Law BA (ed.) *Technology of Cheesemaking*, Sheffield Academic Press, Sheffield, UK, 68–97.
- McSweeney PLH, Ottogalli G, Fox PF (2004):** Diversity of cheese varieties: an overview. In: Fox P.F., McSweeney PLH, Cogan TM, Guinee TP (ed.) *Cheese: Chemistry, physics and microbiology Vol 2 Major cheese groups General aspects*, Elsevier, London, UK, 391–433.
- Mucchetti G, Neviani E (2006):** *Microbiologia e tecnologia lattiero-casearia*. Tecniche Nuove (ed), Milano, IT.
- Mucchetti G, Bonvini B, Remagni MC, Ghiglietti R, Locci F, Barzaghi S, Francolino S, Perrone A, Rubiloni A, Campo P, Gatti M, Carminati D (2008):** Influence of cheese-making technology on composition and microbiological characteristics of Vastedda cheese. *Food Control* 19: 119–125.
- Niro S, Fratianni A, Tremonte P, Sorrentino E, Tipaldi L, Panfili G, Coppola R (2014):** Innovative Caciocavallo cheeses made from a mixture of cow milk with ewe or goat milk. *J Dairy Sci* 97: 1296–1304.
- Núñez M, Gaya P, Medina M, Rodríguez-Marin MA, García-Aser C (1986):** Changes in Microbiological, Chemical, Rheological and Sensory Characteristics during Ripening of Vacuum Packaged Manchego Cheese. *J Food Sci* 51: 1451–1455.
- Palmeri M, Cardamone C., Todaro M, Settanni L, Gaglio R, Mancuso I, Scatassa ML, Corona O (2015):** An investigation of the shelf life of PDO Vastedda della Valle del Belice cheese subjected to different packaging system alternative to vacuum. *Ital J Animal Sci* 14 suppl 1: 43.
- Rudan MA, Barbano DM, Yun JJ, Kindstedt PS (1999):** Effect of fat reduction on chemical composition, proteolysis functionality, and yield of Mozzarella cheese. *J Dairy Sci* 82: 661–672.
- Salari F, Altomonte I, Martini M (2011):** Effetti della stagionatura e del sottovuoto sulle caratteristiche nutrizionali del formaggio Pecorino. *Sci Tecn Latt Cas* 62: 223–229.
- Salvadori Del Prato O (2001):** *Trattato di Tecnologia Casearia*. Calderini-Edagricole, Bologna, IT.
- SAS Institute Inc. (2007):** SAS/STAT Release 9.12. SAS Publishing, Cary, USA.
- Scintu MF Piredda G (2007):** Typicity and biodiversity of goat and sheep milk products. *Small Rumin Res* 68: 221–231.
- Settanni L, Di Grigoli A, Tornambé G, Bellina V, Francesca N, Moschetti G, Bonanno A (2012):** Persistence of wild *Streptococcus thermophilus* strains on wooden vat and during the manufacture of a traditional Caciocavallo type cheese. *Int J Food Microbiol* 155: 73–81.
- Upadhyay VK, McSweeney PLH, Magboul AAA, Fox PF (2004):** Proteolysis in cheese during ripening. In: Fox P.F., McSweeney PLH, Cogan TM, Guinee TP (ed.) *Cheese: Chemistry, physics and microbiology Vol 1 General aspects*, Elsevier, London, UK, 391–433.
- Zannoni M, Annibaldi S (1981):** Standardization of the renneting ability of milk by Formagraph. *Sci Tecn Latt-Cas* 32: 79–94.
- Zottola T, Briganti P, Cuoco E, D’Amici L, De Gregorio A, Guzzon L, Manocchio A, Mancuso M, Condoleo RU (2009):** Studio dei pericoli microbiologici nella produzione della Mozzarella di Bufala Campana. *Abstract Book XIX Convegno AIVI*, 39.

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