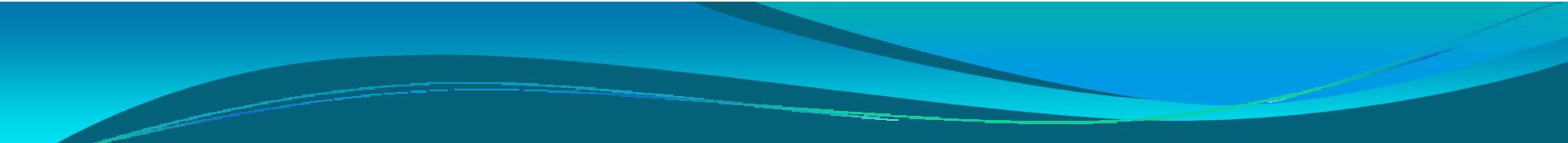


The Molecular Diagnosis of EIA, a Permanent Challenge or an Attainable Goal?

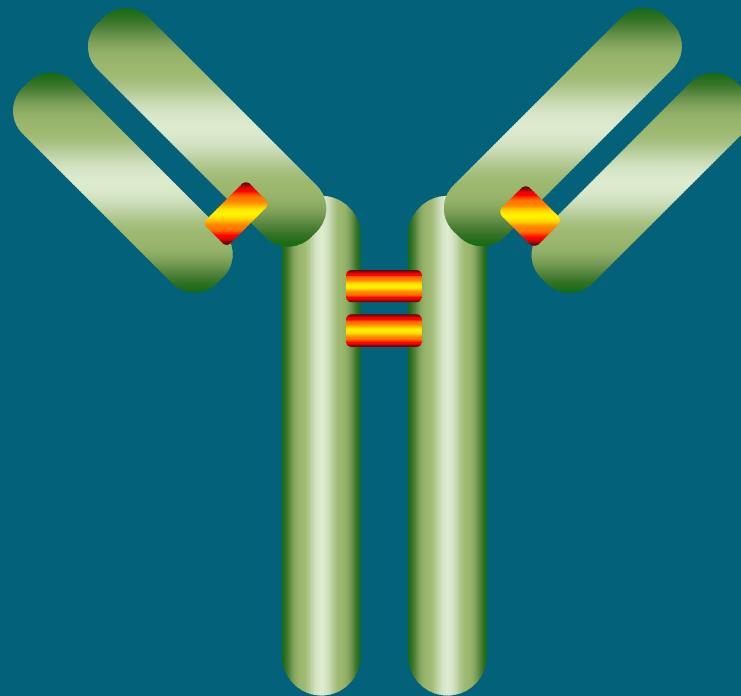


**R. Frank Cook
Gluck Equine Research Center
University of Kentucky**

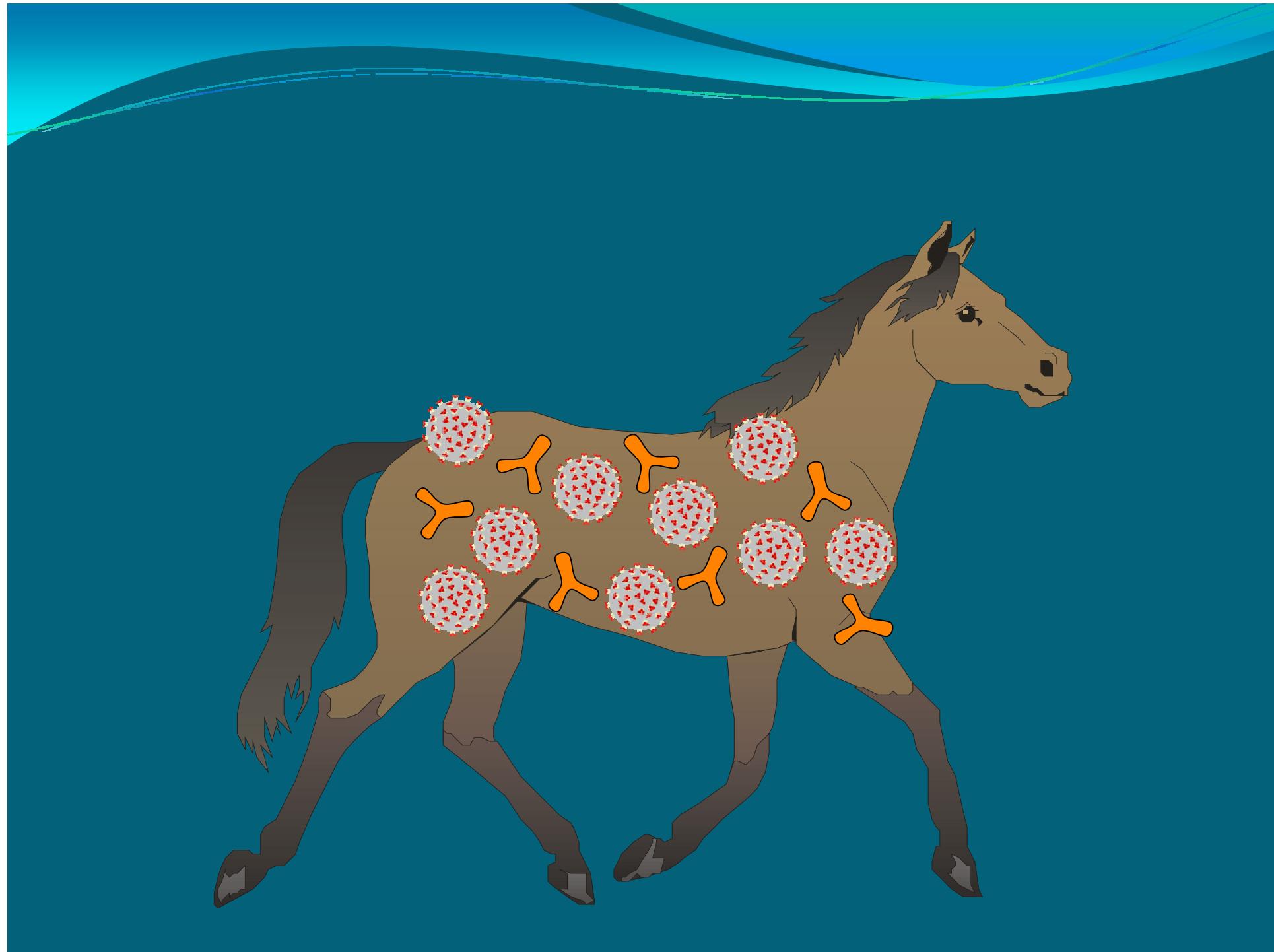


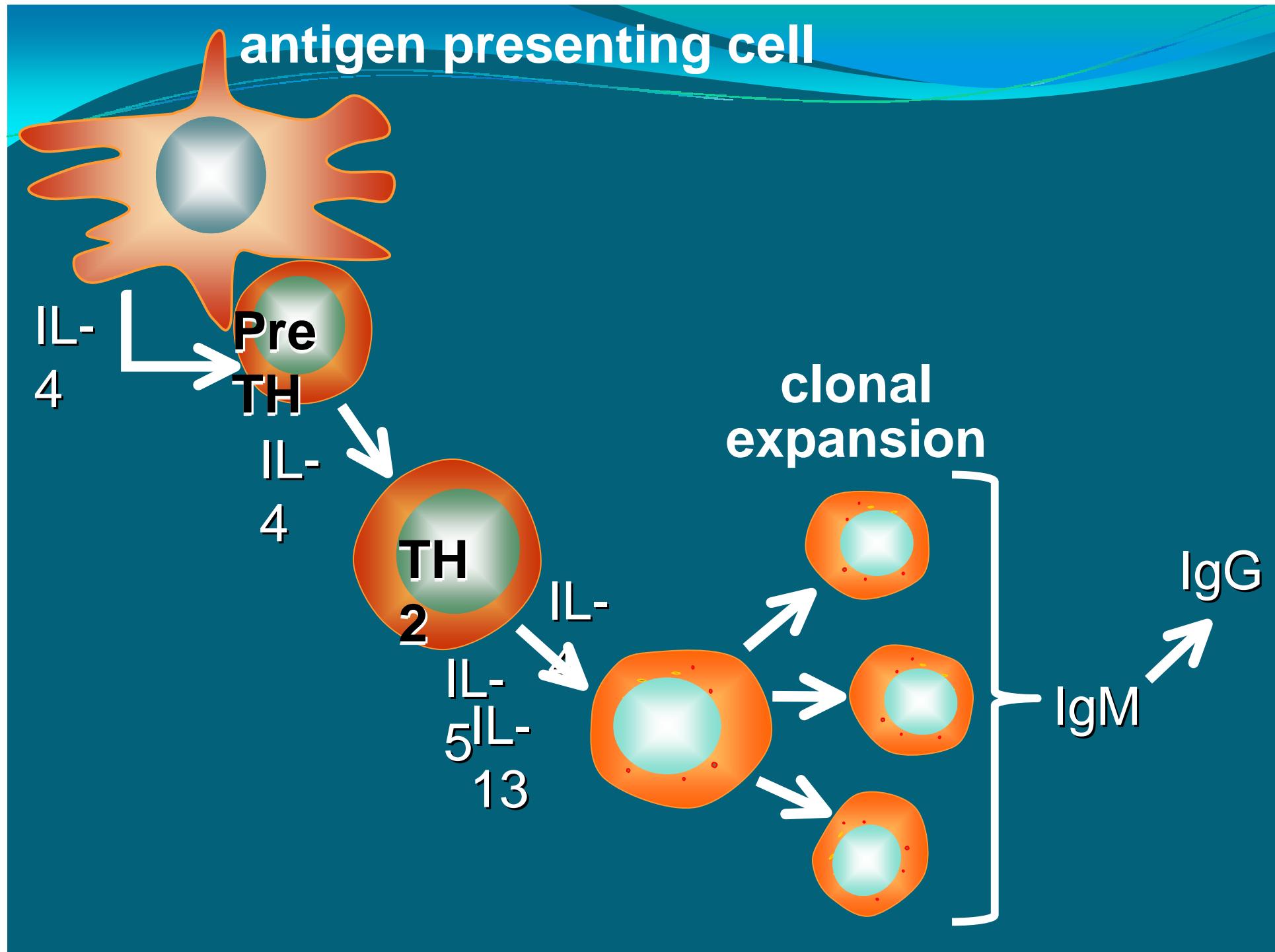
Why is There a Need for Molecular Detection of EIAV?

IAV: Serological Diagnosis



Indirect Method of Detection



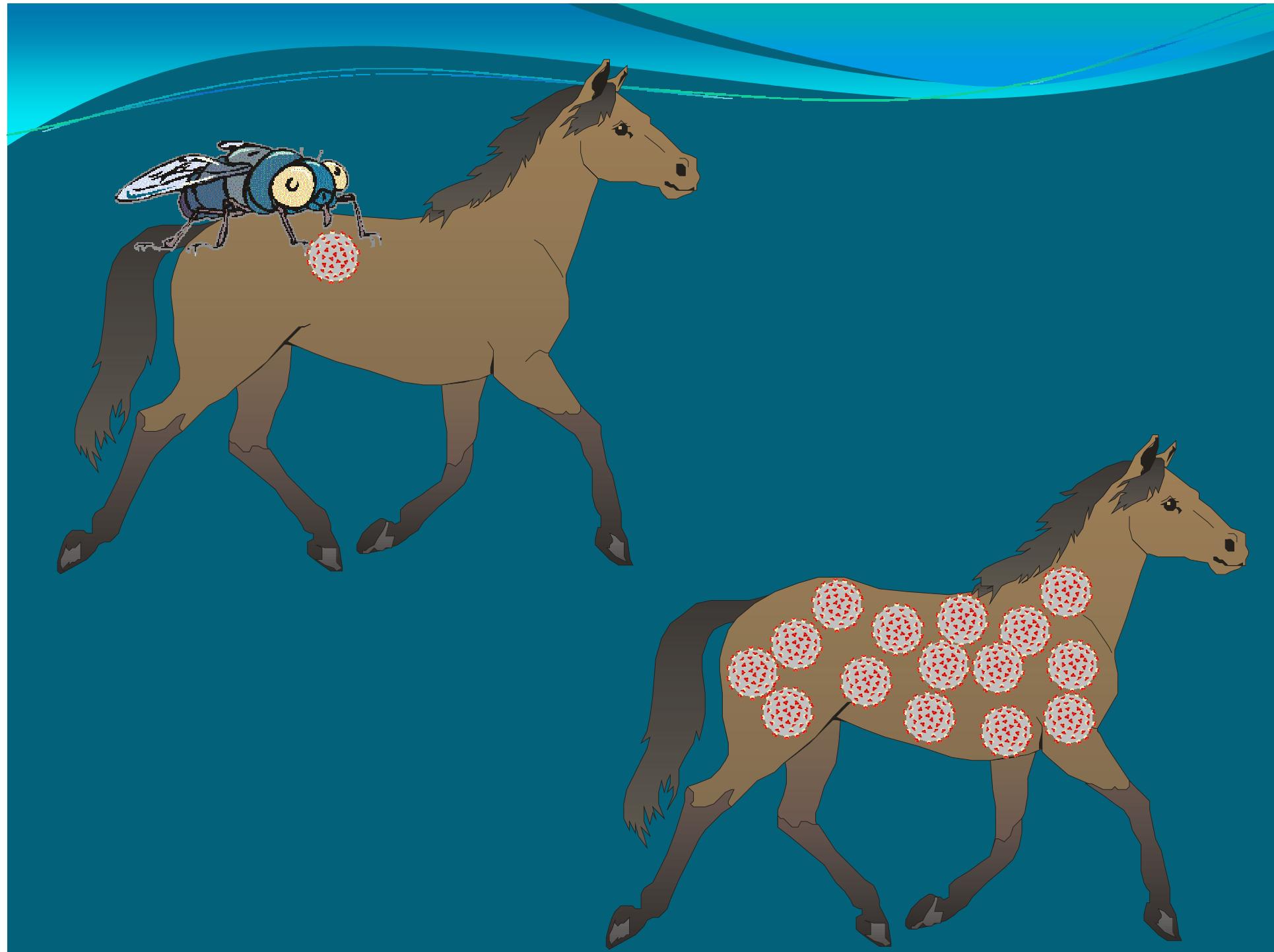


EIA

- Indirect Detection Disadvantages:
 - ❖ Incubation Period (AGID)
 - Most cases within 45 days (Issel & Cook 1993)
 - Longest reported 157 days (Cullinane et. al. 2007)
 - ❖ Italy 2006: Farm
 - 90 day quarantine : ALL AGID –ve
 - 5 months later: 8 NEW cases
 - ❖ Argentina 2012: Riding School
 - Horse with –ve AGID

EIAV : A Need for Direct Detection

Recent
Exposure



EIAV: Direct Detection Methods

Virus Isolation:

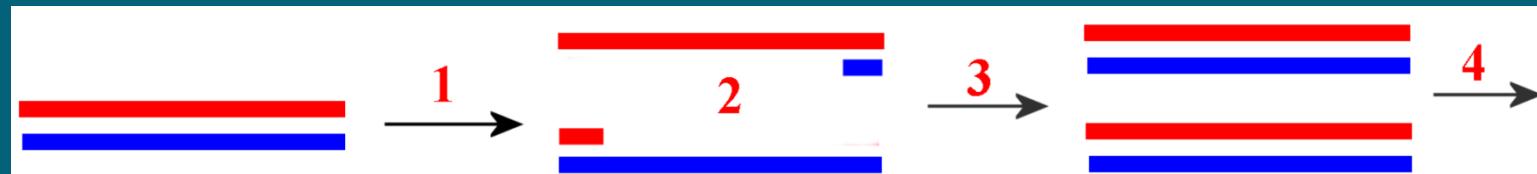
Equine MØ
Insensitive
Highly Variable

Protein: Sensitivity

Nucleic Acid: vRNA/Proviral DNA

PCR
Isothermal

PCR



Cycle #	# DNA Molecules
1	2
2	4
3	8
4	16
5	32
6	64
7	128
8	256
9	512
10	1024

- 1: Denaturation (95°C)**
- 2: Anneal (45-60°C)**
- 3: Elongation (65-75°C)**
- 4: “Repeat 1-3”**

EIAV PCR: Problems

- PCR sensitive to variation in primer/probe binding sites
- EIAV: HIGH Variability
- Inapparent Carrier: LOW viral loads in blood
Undetectable viral RNA

Variation Between EIAV Isolates





PCR Primer Mismatches

Primer TAGGAATTCTCGCC TTAACG

Virus ATCCTTAAGAGCGGAATTGC

Primer TAGGAATTCTCGCC

Virus A CCTTAA AGCGGAATTGC
 A T

annealing

6°C

temp

A/T -2°C G/C -4°C = -

Primer TAGGAATTCTC

GCC

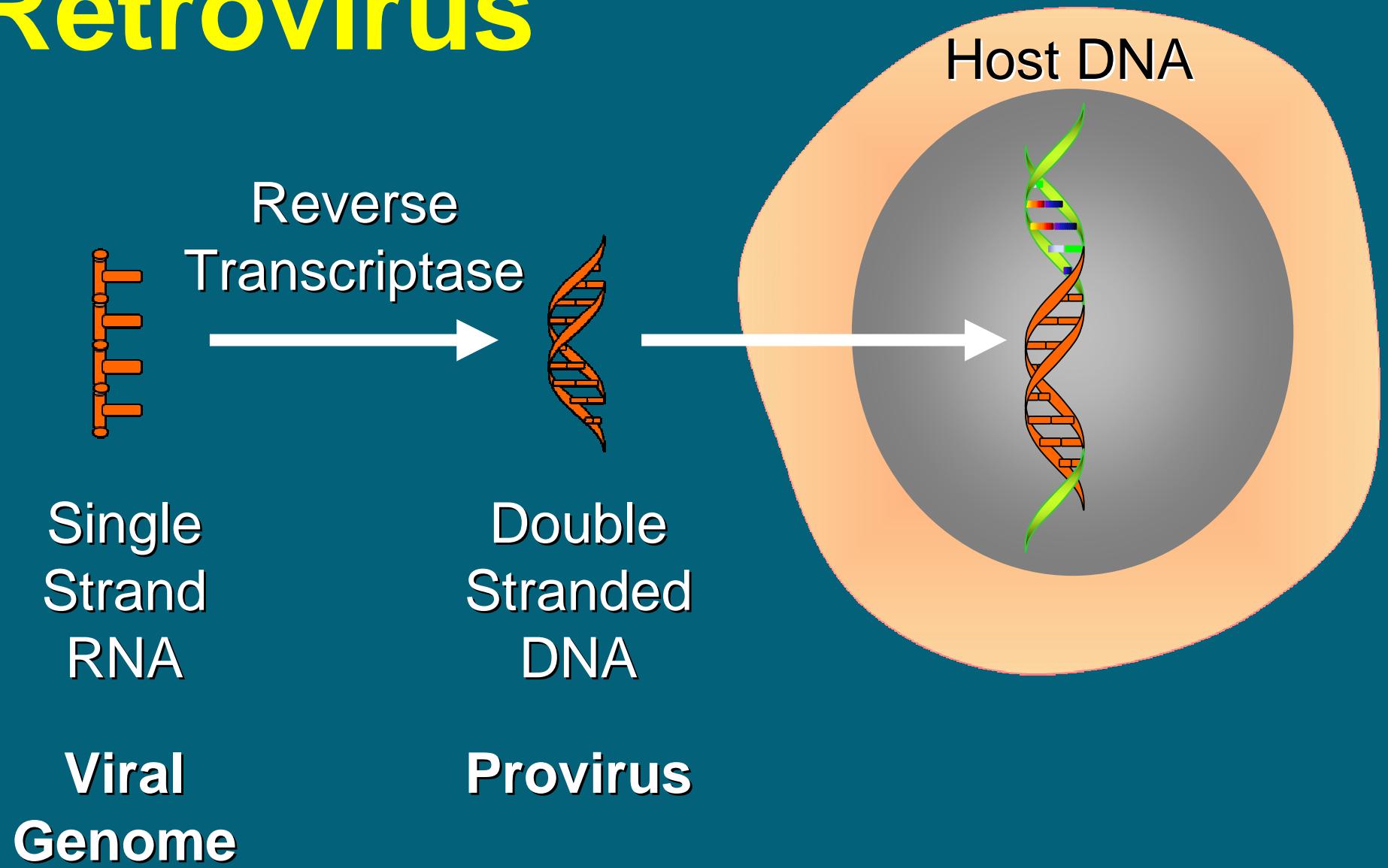
Virus ATCCTTAAGAGAAAAATTGC

annealing temp = 3x G/C = -12°C

EIAV Published PCR Protocols

- Nested
 - ❖ Langemeier et. al. 1996
 - ❖ Nagarajan and Simard, 2001 – OIE
 - ❖ Quinlivan et. al. 2007
 - ❖ Cappelli et. al. 2011
 - ❖ Capomaccio et. al. 2012
 - ❖ Dong et. al. 2012
- Real Time
 - ❖ Cook et.al. 2002

Retrovirus



EIAV: Target Nucleic Acid

- Capelli et. al. 2007

AGID Comparison

❖ vRNA POOR
❖ Proviral DNA GOOD

- Dong et. al. 2012
 - ❖ Proviral DNA
 - ❖ 12/12 EIA Seropositive Horses

Comparative PCR Testing

- Nagarajan and Simard, 2001
- Cappelli et. al., 2011
- Dong et. al., 2012
- Modified Cappelli

Samples: Argentina
26 AGID –ve
25 AGID +ve
3 Febrile

Template: 1µg PBMC DNA

TaKaRa Ex Taq™

Primer Variation

Consensus	1	10	20
1. ITA 1	TG	C . A . G . T . G . . G	
2. ITA 2	TG	C . A . G . T . G . . G	
3. ITA 3	TG	C . A . G . T . G . . G	
4. ITA 5	AT	C . A . G . T . G . . G	
5. IRE	TG	C . A . G . T . G . . G	
6. FL	AC	C . A . G . C . G . . A	
7. NC	AT	C . A . G . T . G . . A	
8. PA	AT	T . A . G . C . G . . A	
9. WY	AT	C . A . G . C . A . . A	
10. Can 1	AT	C . A . G . C . A . . G	
11. Can 3	AT	T . A . G . T . G . . G	
12. Can 7	AC	C . A . A . T . G . . G	
13. Can 10	AT	C . A . G . C . A . . G	
14. Brazil 77	AT	T . A . G . T . G . . G	
15. Brazil 95	AT	T . A . G . T . G . . G	
16. China	AT	C . G . A . T . G . . G	

PCR Results

	NS	C	D	MC
AGID/Blot -ve p26 (26)	0	0	0	0
AGID/Blot +ve p26 (25)	0	3 (12%)	4 (16%)	4 (16%)
FEBRILE (3)	0	3 (21%)	3 (25%)	3 (25%)

NS: Nagarajan and Simard

C: Cappelli et.al.

D: Dong et. al.

MC: Modified Cappelli

PCR +ve Samples

- Cappelli et. al. 3, 20, 31
- Dong et. al. 3, 8, 20, 31
- Modified 8, 12, 20, 31

Conclusions

- PCR Primers reactive against unknown field strains
- N&S – OIE too strain specific?
- Cappelli, Dong, Modified: HIGH specificity; VERY LOW reactivity against endogenous retroviral elements
- LOW sensitivity: Primer mismatches?
Detection limits??

STILL WORK TO DO